

## Full list of publications

(March 09, 2020)

Of the 155 publications, 115 are found in the web of knowledge. *h*-index = 40

Theses (3)

Original Publications (98)

Reviews (42)

Miscellaneous (12)

### Theses (3)

Shima, S. (1983) Effect of ammonium ion on the nitrogenase activity of *Azolla*, Bachelor thesis, Osaka Prefecture University, Osaka, Japan.

Shima, S. (1985) Microbial ecology of *Bradyrhizobium japonicum* strains, Master thesis, Osaka Prefecture University, Osaka, Japan.

Shima, S. (1991) Genetical and biochemical study on cellulases from mesophilic Clostridia, Ph.D. thesis, University of Tokyo, Tokyo, Japan.

### Original Publications (98) (\*corresponding author)

1. Huang, G., Wagner, T., Demmer, U., Warkentin, E., Ermler, U., & Shima, S.\* (2020) The hydride transfer process in NADP dependent methylene-tetrahydromethanopterin dehydrogenase. *J. Mol. Biol.* doi: 10.1016/j.jmb.2020.01.042.
2. Hemmann, J.L., Wagner, T., Shima, S., & Vorholt, J.A.\* (2019) Methylfuran is a prosthetic group of the formyltransferase/hydrolase complex and shuttles one-carbon units between two active sites. *Proc. Natl. Acad. Sci. USA* 116, 25583-25590.
3. Ilina, Y., Lorent, C., Katz, S., Jeoung, J.H., Shima, S., Horch, M.\* Zebger, I.\* & Dobbek, H.\* (2019) X-ray crystallography and vibrational spectroscopy reveal key determinants of biocatalytic dihydrogen cycling by [NiFe] hydrogenases. *Angew. Chem. Int. Edit.* 58,

18710-18714.

4. Huang, G., Wagner, T., Wodrich, M.D., Ataka, K., Bill, E., Ermler, U., Hu X. & Shima, S.\* (2019) The atomic-resolution crystal structure of activated [Fe]-hydrogenase. *Nat. Catal.* 2, 537-543.
5. Pan, H-J, Huang, G., Wodrich, M.D. Tirani, F.F. Ataka, K., Shima, S.\* & Hu, X.\* (2019) A catalytically active [Mn]-hydrogenase incorporating a non-native metal cofactor. *Nat. Chem.* 11, 669–675.
6. Engilberge, S., Wagner, T., Santoni, G., Breyton, C., Shima, S., Franzetti, B., Riobé, F., Mauryd, O. & Girard, E.\* (2019) Protein crystal structure determination with the crystallophore, a nucleating and phasing agent. *J. Appl. Crystallogr.* 52: 722-731.
7. Vögeli, B., Shima, S., Erb, T.\* & Wagner, T.\* (2019) Crystal structure of archaeal HMG-CoA reductase: insights into structural changes of the C-terminal helix of the class-I enzyme. *FEBS Lett.* 593, 543–553.
8. Watanabe, T., Wagner, T., Huang, G., Kahnt, J., Ataka, K., Ermler, U. & Shima, S.\* (2019) The bacterial [Fe]-hydrogenase paralog HmdII uses tetrahydrofolate derivatives as substrates. *Angew. Chem. Int. Ed.* 58, 3506 –3510.
9. Bernhardsgrütter, I., Vögeli, B., Wagner, T., Peter, D.M., Cortina, N.S., Kahnt, J., Bange, G., Engilberge, S., Girard, E., Riobé, F., Maury, O., Shima, S., Zarzycki, J. & Erb, T.J.\* (2018) The multicatalytic compartment of propionyl-CoA synthase sequesters a toxic metabolite. *Nat. Chem. Biol.* 14, 1127–1132.
10. Wagner, T., Huang. G., Ermler, U. & Shima, S.\* (2018) How [Fe]-hydrogenase from *Methanothermobacter* is protected against light and oxidative stress. *Angew. Chem. Int. Ed.* 57, 15056-15059.
11. Vögeli, B., Rosenthal, R.G., Stoffel, G.M.M., Wagner, T., Kiefer, P., Cortina, N.S., Shima, S. and Erb, T.J.\* (2018) InhA, the enoyl-thioester reductase from *Mycobacterium tuberculosis* forms a covalent adduct during catalysis. *J. Biol. Chem.* 293, 17200-17207.
12. Engilberge. S., Riobé, F.\*., Wagner, T., Di Pietro, S., Breyton, C., Franzetti, B., Shima, S., Girard, E.\*., Dumont, E. & Maury, O.\* (2018) Unveiling the binding modes of the crystallophore, a terbium-based nucleating and phasing molecular agent for protein crystallography. *Chem. Eur. J.* 24, 9739-9746.
13. Vögeli, B., Engilberge, S., Girard, E., Riobé, F., Maury, O., Erb, T. J., Shima, S. & Wagner, T.\* (2018) The archaeal acetoacetyl-CoA-thiolase/HMG-CoA-synthase complex channels the intermediate via a fused CoA-binding site. *Pros. Natl. Acad. Sci. USA* 115, 3380-3385.
14. Huang, G., Wagner, T., Ermler, U. Bill, E., Ataka, K. & Shima, S.\* (2018) Dioxygen sensitivity of [Fe]-hydrogenase in the presence of reducing substrates. *Angew. Chem. Int. Ed.*

- 57, 4917-4920.
15. Wagner, T., Koch, J., Ermler, U. & Shima, S.\* (2017) Methanogenic heterodisulfide reductase (HdrABC-MvhAGD) uses two noncubane [4Fe-4S] clusters for reduction. *Science* 357, 699–703.
  16. Bai, L., Wagner, T., Xu, T., Hu, X., Ermler, U. & Shima, S.\* (2017) Water-bridged H-bonding network contributes to the catalysis of a SAM-dependent C-methyltransferase HcgC. *Angew. Chem. Int. Ed.*, 56, 10806–0809.
  17. Wagner, T., Wegner, C.-E., Kahnt, J., Ermler, U. & Shima, S.\* (2017) Phylogenetic and structural comparisons of the three types of methyl-coenzyme M reductase from *Methanococcales* and *Methanobacteriales*. *J. Bacteriol.* 357, 699–703.
  18. Rosenthal, R.G., Vögeli, B., Wagner, T., Shima, S. & Erb, T.J.\* (2017) ‘Negative catalysis’ by a conserved threonine prevents self-intoxication of enoyl-thioester reductases. *Nat. Chem. Biol.* 13, 745-749.
  19. Bai, L., Fujishiro, T., Huang, G., Koch, J., Takabayashi, A., Yokono, M., Tanaka, A., Xu, T., Hu, X., Ermler, U. & Shima, S.\* (2017) Towards artificial methanogenesis: biosynthesis of the [Fe]-hydrogenase cofactor and characterization of the semisynthetic hydrogenase. *Faraday Discussion*, 198, 37-58.
  20. Wagner, T., Ermler, U. & Shima, S.\* (2016) The methanogenic CO<sub>2</sub> reducing-and-fixing enzyme is bifunctional and contains 46 [4Fe-4S] clusters. *Science* 354: 114-117.
  21. Wagner, T., Kahnt, J., Ermler, U. & Shima, S.\* (2016) Didehydroaspartate modification in methyl-coenzyme M reductase catalyzing methane formation. *Angew. Chem. Int. Ed.* 55: 10630-10633.
  22. Upadhyay, V., Ceh, K., Tumulka, F., Abele, R., Hoffmann, J., Langer, J., Shima, S.\* & Ermler, U.\* (2016) Molecular characterization of methanogenic N<sup>5</sup>-methyl-tetrahydromethanopterin: coenzyme M methyltransferase. *Biochim. Biophys. Acta.* 1858: 2140-2144.
  23. Fujishiro, T., Bai, L., Xu, T., Xie, X., Schick, M. Kahnt, J., Rother, M., Hu, X., Ermler, U. & Shima, S.\* (2016) Identification of HcgC as SAM-dependent pyridinol methyltransferase in [Fe]-hydrogenase cofactor biosynthesis. *Angew. Chem. Int. Ed.* 55: 9648-9651.
  24. Wagner, T., Ermler, U. & Shima, S.\* (2016) MtrA of the sodium ion pumping methyltransferase binds cobalamin in a unique mode. *Sci. Rep.* 6, 28226. doi: 10.1038/srep28226.
  25. Duin, E.C., Wagner, T., Shima, S., Prakash, D., Cronin, B., Yáñez-Ruiz, D.R., Duval, S., Ruembeli, R., Stemmler, R.T., Thauer, R.K.\* & Kindermann, M.\* (2016) Mode of action uncovered for the specific reduction of methane emissions from ruminants by the small

- molecule 3-nitrooxypropanol. *Proc. Natl. Acad. Sci. USA*, 113, 6172–6177.
26. Shima, S.\*, Chen, D., Xu, X., Wodrich, M.D., Fujishiro, T., Schultz, K.M., Kahnt, J., Ataka, K. & Hu, X.\* (2015) Reconstitution of [Fe]-hydrogenase using model complexes. *Nat. Chem.* 7, 995–1002.
  27. Hidese, R., Ataka, K., Bill, E. & Shima, S.\* (2015) Cu<sup>I</sup> and H<sub>2</sub>O<sub>2</sub> inactivate and Fe<sup>II</sup> inhibits [Fe]-hydrogenase at very low concentrations. *ChemBioChem* 16, 1861–1865.
  28. Fujishiro, T., Ataka, K., Ermler, U. & Shima, S.\* (2015) Towards a functional identification of catalytically inactive [Fe]-hydrogenase paralogs. *FEBS J.* 282, 3412–3423.
  29. Fujishiro, T., Kahnt, J., Ermler, U. & Shima, S.\* (2015) Protein-pyridinol thioester precursor for biosynthesis of the organometallic acyl-iron ligand in [Fe]-hydrogenase cofactor, *Nat. Commun.* 6: 6895, doi:10.1038/ncomms7895.
  30. Kojima, H., Moll, J., Kahnt, J., Fukui, M. & Shima, S.\* (2014) A reversed genetic approach reveals the coenzyme specificity and other catalytic properties of three enzymes putatively involved in anaerobic oxidation of methane with sulfate. *Environ. Microbiol.* 16: 3431–3442.
  31. Fujishiro, T., Ermler U. & Shima, S.\* (2014) A possible iron delivery function of the dinuclear iron center of HcgD in [Fe]-hydrogenase cofactor biosynthesis. *FEBS Lett.* 588: 2789–2793.
  32. Vitt, S., Ma, K., Warkentin, E., Moll, J., Pierik, A.J., Shima, S.\* & Ermler, U.\* (2014) The F<sub>420</sub>-reducing [NiFe]-hydrogenase complex from Methanothermobacter marburgensis, the first X-ray structure of a group 3 family member. *J. Mol. Biol.* 426: 2813–2826.
  33. Kaneko, M.\*, Takano, Y., Chikaraishi, Y., Ogawa, N.O., Asakawa S., Watanabe, T., Shima, S., Krüger, M., Matsushita, M., Kimura, H. & Ohkouchi, N. (2014) Quantitative analysis of coenzyme F430 in environmental samples: a new diagnostic tool for methanogenesis and anaerobic methane oxidation. *Anal Chem.* 86: 3633–3638.
  34. Fujishiro, T., Tamura, H., Schick, M., Kahnt, J., Xie, X., Ermler, U. & Shima, S.\* (2013) Identification of the HcgB enzyme in [Fe]-hydrogenase-cofactor biosynthesis. *Angew. Chem. Int. Ed.* 52, 12555 –12558.
  35. Tamura, H., Salomone-Stagni, M., Fujishiro, T., Warkentin, E., Meyer-Klaucke, W., Ermler, U. & Shima, S.\* (2013) Crystal structures of [Fe]-hydrogenase in complex with inhibitory isocyanides: implications for H<sub>2</sub>-activation site. *Angew. Chem. Int. Ed.* 52, 9656 –9659.
  36. Milucka, J.\*, Widdel, F. & Shima, S. (2013) Immunological detection of enzymes for sulfate reduction in anaerobic methane-oxidizing consortia. *Environ. Microbiol.* 15, 1561–1571.
  37. Takano, Y.\*, Kaneko, M., Kahnt, J., Imachi, H., Shima, S. & Ohkouchi, M. (2013) Detection

- of coenzyme F<sub>430</sub> in deep-sea sediments, a key molecule for biological methanogenesis. *Organic Geochemistry*, 58, 137–140.
38. Mills, D.J., Vitt, S., Strauss, M., Shima, S. & Vonck, J.\* (2013) De novo modeling of the F<sub>420</sub>-reducing [NiFe]-hydrogenase from a methanogenic archaeon by cryo-electron microscopy. *eLife*, 2, e00218.
39. Upadhyay, V., Demmer, U., Warkentin, E., Moll, J., Shima, S.\* & Ermler, U.\* (2012) Structure and catalytic mechanism of a N<sup>5</sup>,N<sup>10</sup>-methenyltetrahydro methanopterin cyclohydrolase. *Biochemistry* 51, 8435–8443.
40. Schick, M., Xie, X., Ataka, K., Kahnt, J., Linne, U. & Shima, S.\* (2012) Biosynthesis of the iron-guanylylpyridinol cofactor of [Fe]-hydrogenase in methanogenic archaea as elucidated by stable-isotope labeling. *J. Am. Chem. Soc.* 134, 3271–3280.
41. Shima, S.\*, Krueger, M., Weinert, T. Demmer, U., Kahnt, J., Thauer, R.K. & Ermler, U.\* (2012) Structure of a methyl-coenzyme M reductase from Black Sea mats that oxidize methane anaerobically. *Nature* 481, 98–101.
42. Shima, S.\*, Schick, M., Kahnt, J., Ataka, K., Steinbach, K. & Linne, U. (2012) Evidence for acyl–iron ligation in the active site of [Fe]-hydrogenase provided by mass spectrometry and infrared spectroscopy. *Dalton Trans.* 41, 767–771.
43. Basen, M., Krüger, M., Milucka, J., Kuever, J., Kahnt, J., Grundmann, O., Meyerdierks, A., Widdel, F. & Shima, S.\* (2011) Bacterial enzymes for dissimilatory sulfate reduction in a marine microbial mat (Black Sea) mediating anaerobic oxidation of methane. *Environ. Microbiol.* 13, 1370–1379.
44. Shima, S.\* & Ataka, K. (2011) Isocyanides inhibit [Fe]-hydrogenase with very high affinity. *FEBS Lett.* 585, 353–356.
45. Shima, S.\*, Vogt, S., Göbel, A. & Bill E. (2010) Iron-chromophore circular dichroism of [Fe]-hydrogenase: the conformational change required for H<sub>2</sub> activation. *Angew. Chem. Int. Ed.* 49, 9917–9921.
46. Salomone-Stagni, M., Stellato, F., C. Matthew Whaley, M., Vogt, S., Morante, S., Shima, S., Rauchfuss, T.B. & Meyer-Klaucke, W.\* (2010) The iron-site structure of [Fe]-hydrogenase and model systems: an X-ray absorption near edge spectroscopy study. *Dalton Trans.* 39, 3057–3064.
47. Ceh, K., Demmer, U., Warkentin, E., Moll, J., Thauer, R.K., Shima, S. & Ermler, U.\* (2009) Structural basis of the hydride transfer mechanism in F<sub>420</sub> dependent methylene-tetrahydromethanopterin dehydrogenase. *Biochemistry* 48, 10098–10105.
48. Hiromoto, T., Warkentin, E., Moll, J., Ermler, U. & Shima, S.\* (2009) The crystal structure of

- an [Fe]-hydrogenase substrate complex reveals the framework for H<sub>2</sub>-activation. *Angew. Chem. Int. Ed.* 48, 6457–6460.
49. Hiromoto, T., Ataka, K., Pilak, O., Vogt, S., Stagni, M.S., Meyer-Klaucke, W., Warkentin, E., Thauer, R.K., Shima, S.\* & Ermler, U.\* (2009) The crystal structure of C176A mutated [Fe]-hydrogenase suggests an acyl-iron ligation in the active site iron complex. *FEBS Lett.* 583, 585–590.
50. Ettwig, K.F., Shima, S., van de Pas-Schoonen, K.T., Kahnt, J., Medema, M., op den Camp, H.J.M. Jetten, M.S.M. & Strous, M.\* (2008) Denitrifying bacteria oxidize methane in the absence of archaea. *Environ. Microbiol.* 10, 3164–3173.
51. Shima, S.\*, Pilak O., Vogt, S., Schick, M., Stagni, M.S., Meyer-Klaucke, W., Warkentin, E., Thauer, R.K., & Ermler, U.\* (2008) The crystal structure of [Fe]-hydrogenase reveals the geometry of the active site. *Science*, 321, 572–575.
52. Mayr, S., Latkoczy C., Krüger, M., Günther, D., Shima, S., Thauer, R.K., Widdel, F. & Jaun, B.\* (2008) The structure of a F<sub>430</sub> variant from archaea associated with anaerobic oxidation of methane. *J. Am. Chem. Soc.*, 130, 10758–10767.
53. Guo, Y., Wang, H., Xiao, Y., Vogt, S., Thauer, R.K.\*, Shima, S., Volkers, P.I., Rauchfuss, T.B.\* Pelmentschikov, V., Case, D.A.\* Alp, E.E., Sturhahn, W., Yoda, Y. & Cramer, S.P.\* (2008) Characterization of the Fe site in the iron-sulfur-cluster-free hydrogenase (Hmd) and of a model compound via nuclear resonance vibrational spectroscopy (NRVS). *Inorg. Chem.* 47, 3969–3977.
54. Vogt, S., Lyon, E.J., Shima, S. & Thauer, R.K.\* (2008) The exchange activities of [Fe]-hydrogenase (iron-sulfurcluster- free hydrogenase) from methanogenic archaea in comparison with the exchange activities of [FeFe] and [NiFe] hydrogenases. *J. Biol. Inorg. Chem.* 13, 97–106.
55. Seedorf, H., Hagemeier, C.H., Shima, S., Thauer, R.K., Warkentin, E. & Ermler, U.\* (2007) Structure of coenzyme F<sub>420</sub>H<sub>2</sub> oxidase (FprA), a di-iron flavoprotein from methanogenic archaea catalyzing the reduction of O<sub>2</sub> to H<sub>2</sub>O. *FEBS J.* 274, 1588–1599.
56. Kahnt, J., Buchenau, B., Mahlert, F., Krüger, M., Shima, S. & Thauer, R.K.\* (2007) Post-translational modifications in the active site region of methyl-coenzyme M reductase from methanogenic and methanotrophic archaea. *FEBS J.* 274, 4913–4921.
57. Korbas, M., Meyer-Klaucke, W.\* Vogt, S., Lyon, E. J. Thauer, R. K. & Shima, S.\* (2006) The iron-sulfur cluster free hydrogenase (Hmd) is a metalloenzyme with a novel iron binding motif. *J. Biol. Chem.* 281, 30804–30813.
58. Pilak, O., Mamat, B., Vogt, S., Hagemeier, C.H., Thauer, R.K., Shima, S.\*, Vonrhein, C., Warkentin, E. & Ermler, U.\* (2006) The crystal structure of the apoenzyme of the iron-sulfur-

- cluster-free hydrogenase (Hmd). *J. Mol. Biol.* 358: 798-809.
59. Acharya, P., Warkentin, E., Ermler, U.\*, Thauer, R.K. & Shima, S.\* (2006) The structure of formylmethanofuran:tetrahydromethanopterin formyltransferase in complex with its coenzymes. *J. Mol. Biol.* 357: 870–879.
60. Shima, S.\*, Lyon, E. J., Thauer, R. K. Mienert, B. & Bill, E.\* (2005) Mössbauer studies of the iron-sulfur-cluster-free Hydrogenase (Hmd): The electronic state of the mononuclear Fe active site. *J. Am. Chem. Soc.* 127, 10430–10435.
61. Aufhammer, S. W., Warkentin, E., Ermler, U., Hagemeier, C. H., Thauer, R. K. & Shima, S.\* (2005) Crystal structure of methylenetetrahydro- methanopterin reductase (Mer) in complex with coenzyme F<sub>420</sub>: Architecture of the F<sub>420</sub>/FMN binding site of enzymes within the non-prolyl *cis*-peptide containing bacterial luciferase family. *Protein Sci.* 14, 1840–1849.
62. Warkentin, E.\*, Hagemeier, C.H., Shima, S., Thauer R.K. & Ermler, U. (2005) The structure of F<sub>420</sub>-dependent methylenetetrahydro- methanopterin dehydrogenase: a crystallographic superstructure of the selenomethionine-labelled protein crystal structure. *Acta Crystallog. Sect. D.* 61: 198–202.
63. Sakasegawa, S., Hagemeier, S. C., Thauer R. K. Essen, L.-O.\* & Shima, S.\* (2004) Structural and functional analysis of the *gpsA* gene product of *Archaeoglobus fulgidus*: A glycerol-3-phosphate dehydrogenase with unusual NADP<sup>+</sup> preference. *Protein Sci.* 13, 3162-3171.
64. Lyon, E. J., Shima, S., Boecker, R., Thauer, R. K.\*, Grevels, F.-W., Bill, E., Roseboom, W. Albracht, S. P. J.\* (2004) Carbon monoxide as an intrinsic ligand to iron in the active site of the iron-sulfur cluster free hydrogenase (Hmd) as revealed by infrared spectroscopy. *J. Am. Chem. Soc.* 126, 14239–14248.
65. Seedorf, H., Dreisbach, A., Hedderich, R., Shima, S. & Thauer, R.K.\* (2004) F<sub>420</sub>H<sub>2</sub>-oxidase (FprA) from *Methanobrevibacter arboriphilus*, a novel coenzyme F<sub>420</sub> dependent enzyme involved in O<sub>2</sub> detoxification. *Arch. Microbiol.* 182, 126–137.
66. Shima, S., Lyon, E. J., Sordel-Klippert, Kauß, M., Kahnt, J., Thauer, R. K.\*., Steinbach, K., Xie, X., Verdier, L. & Griesinger, C.\* (2004) The cofactor of the iron-sulfur cluster free hydrogenase Hmd: structure of the light-inactivation product. *Angew. Chemie Int. Ed.* 43, 2547–2551.
67. Aufhammer, S., Warkentin, E., Berk, H., Shima, S., Thauer, R.K. & Ermler, U.\* (2004) Coenzyme binding in F<sub>420</sub>-dependent alcohol dehydrogenase, a member of the bacterial luciferase family. *Structure* 12, 361–370.
68. Lyon, E.J., Shima, S., Buurman, G., Chowdhuri, S., Batschauer, A., Steinbach, K., Thauer, R.K.\* (2004) UV-A/blue light inactivation of the “metal-free” hydrogenase (Hmd) from methanogenic archaea: the enzyme appears to contain functional iron after all. *Eur. J.*

- Biochem.* 271, 195–204.
69. Krüger, M., Meyerdierks, A., Glöckner, F.O., Amann, R., Widdel, F.\*, Kube, M., Reinhardt, R., Kahnt, J., Böcher, R., Thauer, R.K.\* & Shima, S. (2003) An conspicuous nickel protein in microbial mats that oxidize methane anaerobically. *Nature* 426, 878–881.
70. Hagemeier, C.H., Shima, S., Thauer, R.K., Bourenkov, G., Bartunik, H.D. & Ermler, U.\* (2003) Coenzyme F<sub>420</sub>-dependent methylenetetrahydromethanopterin dehydrogenase (Mtd) from *Methanopyrus*: A methanogenic enzyme with an unusual quarternary structure. *J. Mol. Biol.* 332, 1047–1057.
71. Hagemeier, C.H., Shima, S., Warkentin, E., Thauer, R.K. & Ermler, U.\* (2003) Coenzyme F<sub>420</sub> dependent methylenetetrahydromethanopterin dehydrogenase from *Methanopyrus kandleri*: the selenomethionine- labelled and non-labelled enzyme crystallized in two different forms. *Acta Crystallogr. Sect. D* 59, 1635–1655.
72. Mamat, B., Roth, A., Grimm,, C., Ermler, U., Tziatzios, C., Schubert, D., Thauer, R.K. & Shima, S.\* (2002) Crystal structures and enzymatic properties of three formyltransferases from archaea: Environmental adaptation and evolutionary relationship. *Protein Sci.* 11, 2168–2178.
73. Warkentin, E., Mamat, B., Sordel-Klippert, M., Wicke, M., Thauer, R.K., Iwata, M., Iwata, S.\*., Ermler, U.\* & Shima, S.\* (2001) Structures of F<sub>420</sub>H<sub>2</sub>:NADP<sup>+</sup> oxidoreductase with and without its substrates bound. *EMBO J.* 20, 6561–6569.
74. Shima, S.\*, Sordel-Klippert, M., Brioukhanov, A., Netrusov, A., Linder, D. & Thauer, R. K. (2001) Characterization of heme-dependent catalase from *Methanobrevibacter arboriphilus*. *Appl. Environ. Microbiol.* 67, 3041–3045.
75. Grabarse, W., Mahlert, F., Duin, E. C., Goubeaud, M., Shima, S., Thauer, R. K., Lamzin, V. & Ermler, U.\* (2001) On the mechanism of biological methane formation: Structural evidence for conformational changes in methyl-coenzyme M reductase upon substrate binding. *J. Mol. Biol.* 309, 315–330.
76. Buurman, G., Shima, S. & Thauer, R.K.\* (2000) The metal-free hydrogenase from methanogenic archaea: Evidence for a bound cofactor. *FEBS Lett.* 485, 200–204.
77. Shima, S.\*, Thauer, R.K., Ermler, U., Durchschlag, H., Tziatzios, C. & Schubert, D. (2000) A mutation affecting the association equilibrium of formyltransferase from the hyperthermophilic *Methanopyrus kandleri* and its influence on the enzyme's activity and thermostability. *Eur. J. Biochem.* 267, 6619–6623.
78. Grabarse, W., Mahlert, F., Shima, S., Thauer, R.K. & Ermler, U.\* (2000) Comparison of three methyl coenzyme M reductase from phylogenetically distant organisms: Unusual amino acid modification, conservation and adaptation. *J. Mol. Biol.* 303, 329–344.

79. Brioukhanov, A., Netrusov, A., Sordel, M., Thauer, R.K. & Shima, S.\* (2000) Protection of *Methanoscincina barkeri* against oxidative stress: identification and characterization of an iron superoxide dismutase. *Arch. Microbiol.* 174, 213–216.
80. Shima, S., Warkentin, E., Grabarse, W., Sordel, M., Wicke, M., Thauer, R.K. & Ermler, U.\* (2000) Structure of coenzyme F<sub>420</sub> dependent methylenetetrahydromethanopterin reductase from two methanogenic archaea. *J. Mol. Biol.* 300, 935–950.
81. Selmer, T., Kahnt, J., Goubeaud, M., Shima, S., Grabarse, W., Ermler, U & Thauer, R.K.\* (2000) On the biosynthesis of methylated aminoacids in the active site region of methyl-coenzyme M reductase. *J. Biol. Chem.* 275, 3755–3760.
82. Grabarse, W., Vaupel, M., Vorholt, J.A., Shima, S., Thauer, R.K. Wittershagen, A., Bourenkov, G., Bartunik, H.D. & Ermler, U.\* (1999) The crystal structure of methenyltetrahydromethanopterin cyclohydrolase from the hyperthermophilic archaeon *Methanopyrus kandleri*. *Struct. Fold. Des.* 7, 1257–1268.
83. Shima, S.\*, Netrusov, A., Sordel, M., Wicke, M., Hartmann, G.C. & Thauer, R.K. (1999) Purification, characterization, and primary structure of a monofunctional catalase from *Methanoscincina barkeri*. *Arch. Microbiol.* 171, 317–323.
84. Shima, S.\*, Tziatzios, C., Schubert, D., Fukada, H., Takahashi, K., Ermler, U. & Thauer, R.K. (1998) Lyotropic-salt-induced changes in monomer/dimer/tetramer association equilibrium of formyltransferase from the hyperthermophilic *Methanopyrus kandleri* in relation to the activity and thermostability of the enzyme. *Eur. J. Biochem.* 258, 85–92.
85. Shima, S., Héault D.A. Berkessel, A. & Thauer, R.K.\* (1998) Activation and thermostabilization effects of cyclic 2,3-diphosphoglycerate on the enzymes from the hyperthermophilic *Methanopyrus kandleri*. *Arch. Microbiol.* 170, 469–472.
86. Ermler, U.\*, Grabarse, W., Shima, S., Goubeaud, M. & Thauer, R.K. (1997) Crystal structure of methyl-coenzyme M reductase: the key enzyme of biological methane formation. *Science* 278, 1457–1462.
87. Shima, S.\*, Goubeaud, M., Vinzenz, D., Thauer, R.K. & Ermler, U.\* (1997) Crystallization and preliminary X-ray diffraction studies of methyl-coenzyme M reductase from *Methanobacterium thermoautotrophicum*. *J. Biochem. (Tokyo)* 121, 829–830.
88. Ermler, U.\*, Merckel, M.C., Thauer, R.K. & Shima, S. (1997) Formylmethanofuran:tetrahydromethanopterin formyltransferase from *Methanopyrus kandleri*-new insights into salt-dependence and thermostability. *Structure* 5, 635–646.
89. Shima, S., Thauer, R.K., Michel, H. & Ermler, U.\* (1996) Crystallization and preliminary X-ray diffraction studies of formylmethanofuran: tetrahydromethanopterin formyltransferase

- from *Methanopyrus kandleri*. *Proteins: Structure, Function, and Genetics* 26, 118–120.
90. Kunow, J.\*, Shima, S., Vorholt, J. & Thauer, R.K. (1996) Primary structure and properties of the formyltransferase from the mesophilic *Methanosarcina barkeri*: Comparison with the enzymes from thermophilic and hyperthermophilic methanogens. *Arch. Microbiol.* 165, 97–105.
  91. Shima, S., Weiss, D. & Thauer, R.K.\* (1995) Formylmethanofuran: tetrahydromethanopterin formyltransferase (Ftr) from the hyperthermophilic *Methanopyrus kandleri*: Cloning, sequencing and functional expression of the *ftr* gene and one step purification of the enzyme overproduced in *Escherichia coli*. *Eur. J. Biochem.* 230, 906–913.
  92. Shima, S.\*, Yanagi, M. & H. Saiki (1994) The phylogenetic position of *Hydrogenobacter acidophilus* based on 16S rRNA sequence analysis. *FEMS Microbiol. Lett.* 119, 119–122.
  93. Shima, S.\* & Suzuki, K. (1993) *Hydrogenobacter acidophilus* sp. nov., a thermophilic, aerobic, hydrogen-oxidizing bacterium requiring elemental sulfur for growth. *Int. J. Syst. Bacteriol.* 43, 703–708.
  94. Shima, S., Igarashi, Y. & Kodama, T.\* (1993) Purification and properties of two truncated endoglucanases produced in *Escherichia coli* harbouring *Clostridium cellulolyticum* endoglucanase gene *celCCD*. *Appl. Microbiol. Biotechnol.* 38, 750–754.
  95. Shima, S., Igarashi, Y. & Kodama, T.\* (1991) Nucleotide sequence analysis of the endoglucanase-encoding gene, *celCCD*, of *Clostridium cellulolyticum*. *Gene* 104, 33–38.
  96. Shima, S., Igarashi, Y. & Kodama, T.\* (1991) Molecular cloning of a new endoglucanase gene from *Clostridium cellulolyticum* and its expression in *Escherichia coli*. *Appl. Microbiol. Biotechnol.* 35, 233–236.
  97. Shima, S., Kato, J., Igarashi, Y. & Kodama, T.\* (1989) Cloning and expression of a *Clostridium cellobioparum* cellulase gene and its excretion from *Escherichia coli* JM109. *J. Ferment. Bioeng.* 68, 75–78.
  98. Ozawa T.\*, Shima, S. & Yamaguchi, M. (1988) Soil aggregate as a favorable habitat for *Bradyrhizobium japonicum* strains. *Soil Sci. Plant Nutr.* 34, 605–608.

## Reviews (40)

1. Shima, S.\*, Huang, G., Wagner, T., & Ermler, U. Insights from structural biology of hydrogenotrophic methanogenesis. *Annu. Rev. Microbiol.* Submitted

2. Huang, G., Wagner, T., Ermler, U., & Shima, S.\* (2020) Methanogenesis involves direct hydride transfer from H<sub>2</sub> to an organic substrate. *Nat. Rev. Chem.* <https://doi.org/10.1038/s41570-020-0167-2>.
3. Wagner, T., Ermler, U. & Shima, S.\* (2018) Tungsten-containing formylmethanofuran dehydrogenase. In *Encyclopedia of Inorganic and Bioinorganic Chemistry* (online). (A. Messerschmidt, Albrecht ed.) John Wiley and Sons, Inc..
4. Wagner, T., Watanabe, T. & Shima, S.\* (2018) Hydrogenotrophic methanogenesis. In *Handbook of Hydrocarbon and Lipid Microbiology Series. Biogenesis of Hydrocarbons* (A.J.M. Stams and D.Z. Sousa eds.) Springer, Germany.
5. Shima, S. (2017) Methanogenic metabolisms. In *Archaea Biology* (Y. Ishino and H. Atomi Eds) Kyoritsu-Shuppan (in Japanese).
6. Scheller, S., Ermler, U. & Shima, S. (2017) Catabolic pathways & enzymes involved in the anaerobic oxidation of methane. In 6-volume *Handbook of Hydrocarbon and Lipid Microbiology* (M. Boll ed.) Springer, Germany.
7. Shima, S. & Fujishiro, T. (2016) Identification of the biosynthetic enzymes of the [Fe]-hydrogenase cofactor based on the crystal structure of the proteins. *Kouso-kogaku News* 76: 26-30 (in Japanese).
8. Shima, S (2016) [Highlight] The biological methane-forming reaction: Mechanism confirmed through spectroscopic characterization of key intermediate. *Angew. Chem. Int. Ed.* 55:13648-13649.
9. Shima, S. (2015) An energy metabolism under anoxic conditions: Enzyme chemistry of methanogenesis. *Kagakutokogyo* 68, 706-708 (in Japanese).
10. Shima, S., Fujishiro, T. & Ermler, U. (2015) Structure and function of [Fe]-hydrogenase and biosynthesis of the FeGP cofactor. In *Biohydrogen* (M. Roegner ed.) DE Gruyter, Berlin, Germany.
11. Fujishiro, T. & Shima, S. (2014) Functional analysis based on tertiary structure of proteins. *Seibutsukougakukaishi* 92, 676 (in Japanese).
12. Shima, S. (2014) Life under anaerobic conditions: Enzymes involved in hydrogenotrophic methanogenesis. In *Biomolecules under Extreme Environments*. (The Chemical Society of Japan, ed) Kagakudojin, Kyoto, Japan (in Japanese).
13. Shima, S. (2014) Enzyme chemistry of methanogenesis and anaerobic oxidation of methane. *Kagakutoseibutsu* 52, 307–312 (in Japanese).

14. Shima, S. & Ermler, U. (2012) Crystal structure and biochemistry of methyl-coenzyme M reductase from Black Sea mats mediating anaerobic oxidation of methane. *Biseibutsu Seitai* 27, 55–62 (in Japanese).
15. Shima, S. & Ermler U. (2011) Structure and function of [Fe]-hydrogenase and its iron-guanylylpyridinol (FeGP) cofactor. *Eur. J. Inorg. Chem.* 2011, 963–972.
16. Shima, S., Schick, M. & Tamura, H. (2011) Preparation of [Fe]-hydrogenase from methanogenic archaea. *Methods Enzymol.* 494, 119–137.
17. Thauer, R.K., Kaster, A.-K., Goenrich, M., Schick, M., Hiromoto, T. & Shima, S. (2010) Hydrogenases from methanogenic archaea, nickel, a novel cofactor and H<sub>2</sub>-storage. *Ann. Rev. Biochem.* 79: 507–536.
18. Shima, S., Thauer, R.K. & Ermler, U. (2009) Carbon monoxide as intrinsic ligands to iron in the active site of [Fe]-hydrogenase. In *Metal-carbon bonds in enzymes and cofactors*, Vol. 6 of *Metal Ions in Life Sciences* (Sigel, A., Sigel, H., Sigel, R.K.O., eds). John Wiley & Sons, Ltd, Chichester, UK, pp 219–240.
19. Shima, S. (2008) The structure of the [Fe]-hydrogenase and the convergent evolution of the active site of hydrogenases. *Seikagaku* 80, 846–849 (in Japanese).
20. Shima, S. (2008) Functions of methyl-coenzyme M reductase in production and degradation of methane, pp. 182–183. In *Applied Microbiology* (Kumagai, H., Kato, N., Murata, K. & Sakai, Y., eds) Asakura Shoten, Tokyo, Japan (in Japanese).
21. Thauer, R.K. & Shima, S. (2008) Methane as fuel for anaerobic microorganisms. *Ann. NY Acad. Sci.* 1125: 158–170.
22. Shima, S. & Thauer, R.K. (2007) A third type of hydrogenase catalyzing H<sub>2</sub> activation. *Chem. Rec.* 7, 37–46.
23. Thauer R.K. & Shima, S. (2007) Methyl-coenzyme M reductase in methanogenic and methanotrophic archaea. In *Archaea* (Garrett, R. & Klenk, H.-P., eds) Blackwell Publishing, Inc. Malden, USA, pp 275–283.
24. Thauer R.K. & Shima, S. (2006) Methane and microbes. *Nature* 440: 878–879.
25. Thauer R.K. & Shima, S. (2006) Methyl-coenzyme M reductase in methanogenic and methanotrophic archaea. In *Archaea Biology* (Garrett, R. & Klenk, H.-P., eds) Blackwell Publishing, Inc. Malden, USA.
26. Shima, S. & Thauer, R.K. (2006) Anaerobic methane oxidation by archaea: a biochemical approach. *Bioscience and Industry* 64: 23–26 (in Japanese).

27. Shima, S. & Thauer, R.K. (2005) Methyl-coenzyme M reductase (MCR) and the anaerobic oxidation of methane (AOM) in methanotrophic archaea. *Curr. Opin. Microbiol.* 8, 643–648.
28. Shima, S., Thauer, R.K. & Ermler, U. (2004) Hyperthermophilic and salt-dependent formyltransferase from *Methanopyrus kandleri*. *Biochem. Soc. Trans.* 32, 269–272.
29. Shima, S., Warkentin, E., Thauer, R.K. & Ermler, U. (2002) Structure and function of enzymes involved in the methanogenic pathway utilizing carbon dioxide and molecular hydrogen. *J. Biosci. Bioeng.* 93, 519–530.
30. Shima, S. (2002) Energy metabolism of methanogens, pp. 325-333. in Great development of microorganisms (Imanaka, T., Ed.). NTS inc., Tokyo (in Japanese).
31. Grabarse, W., Shima, S., Mahlert, F., Duin, E.C. & Thauer, R.K. & Ermler, U. (2001) Methyl-coenzyme M reductase, pp. 897–914. In *Handbook of Metalloproteins* (Wieghardt, K., Huber, R., Poulos, T.L., Messerschmidt, A.). John Wiley & sons.
32. Shima, S. & Thauer R.K. (2001) Tetrahydromethanopterin specific enzymes from *Methanopyrus kandleri*. *Methods Enzymol.* 331, 317–353.
33. Ermler, U., Grabarse, W., Shima, S., Goubeaud, M. & Thauer, R.K. (1998) Active sites of transition metal enzymes with focus on nickel. *Curr. Opin. Struct. Biol.* 8 749–758.
34. Shima, S. (1998) Mechanisms of methane formation, pp. 190-205. In Y. Koga, and M. Kamekura (eds), Biology of the Archaebacteria. University of Tokyo Press (in Japanese).
35. Shima, S. (1998) Unique structures of the enzymes from methanogens. *Seibutsu-Kogaku Kaishi* 76, 353 (in Japanese).
36. Shima, S. (1998) Mechanism of biological methane formation: Structure and function of methyl-coenzyme M reductase. *Protein, Nucleic acid and Enzyme* 43, 1461–1467 (in Japanese).
37. Ermler, U., Grabarse, W., Shima, S., Goubeaud, M. & Thauer, R.K. (1998) Mechanismus der mikrobiellen Methanbildung. *Biospektrum* 4, 20–24 (in German).
38. Suzuki, K. & Shima, S. (1993) *Hydrogenobacter acidophilus*: Is its ancestor a missing-link between the Bacteria and the Archaea? *RIKEN Review* 3, 3–4 (in Japanese).
39. Shima, S. (1993) Cellulases of clostridia: genetics and biochemistry. *The Heredity* 47, 56–60 (in Japanese).

40. Shima, S. (1990) Application of energy-conversion enzymes for hydrogen production and electrodes of fuel cells. *Energy forum* No. 432, 112 (in Japanese).
41. Shima, S. (1990) Production of organic materials by electric energy: Utilization of hydrogen bacteria. *Nogyodenka* 43, 13–16 (in Japanese).
42. Shima, S. (1985) Utilization of root-nodule bacteria and its ecological problems. *Nogyodenka* 38, 15–17 (in Japanese).

### Miscellaneous (12)

1. Shima, S. (2018) Artificial photosynthesis and methanogenesis (2018) **CanApple News**. On the website of CanApple (in Japanese).
2. Wagner, T., Ermler, U. & Shima, S. (2018) *Japanese Scientist in Science* 2016. 2017, 53.
3. Artero, V., Hammarström, L., Fan, F., Whang, D.R., Martinez, J., Harriman, A., Noguchi, T., Karlsson, J., Summers, P., Itoh, S., Cogdell, R., Kibler, A., Ehrmaier, J., Tamiaki, H., Fujita, E., Shima, S., Yoshino, S., Inoue, H., Wasielewski, M., Corry, T., Gust, D., Cassiola, F., Ishida, H., Takagi, K., Kang, S.O., Li, C., Sun, L., Park, H., Hashimoto, H., Amao, Y., Son, E.J., Kamiya, N., Shen, J.R. & Yamaguchi, K. (2017) Biological approaches to artificial photosynthesis, fundamental processes and theoretical approaches: general discussion. *Faraday Discussion* 198: 147-168 (Discussion part).
4. Wang, M., Artero, V., Hammarström, L., Martinez, J., Karlsson, J., Gust, D., Summers, P., Machan, C., Brueggeller, P., Windle, C.D., Kageshima, Y., Cogdell, R., Tolod, K.R., Kibler, A., Apaydin, D.H., Fujita, E., Ehrmaier, J., Shima, S., Gibson, E., Karadas, F., Harriman, A., Inoue, H., Kudo, A., Takayama, T., Wasielewski, M., Cassiola, F., Yagi, M., Ishida, H., Franco, F., Kang, S.O., Nocera, D., Li, C., Di Fonzo, F., Park, H., Sun, L., Setoyama, T., Kang, Y.S., Ishitani, O., Shen, J.R., Son, H.J., Masaoka, S. (2017) Molecular catalysts for artificial photosynthesis: general discussion. *Faraday discussion* 198:353-395. (Discussion part)
5. Wagner, T., Ermler, U. & Shima, S. (2017) *Japanese Scientist in Science* 2017. 2018.
6. Shima, S. (2011) Application of the function of [Fe]-hydrogenase. *Kagaku Keizai* 58, 93–94 (in Japanese).

Reports of Central Research Institute of Electric Power Industry  
(in Japanese with English abstract)

1. Shima, S. (1993) Purification of the cytoplasmic hydrogenase of thermoacidophilic aerobic hydrogen-oxidizing bacterium, strain 3H-1, and characterization of the strain. (Research

Report U92048).

2. Shima, S. (1991) Isolation of hydrogen bacteria growing in extreme environments and characterization of their hydrogenases. (Research Report: U91009).
3. Shima, S., Watanabe, Y., Saiki, H. & Kiyono, M. (1990) Microbial CO<sub>2</sub> fixation -1- its effect on total emission of greenhouse effect gases. (Research Report U90020).
4. Shima, S. (1989) Molecular breeding of hyper-hydrogen-producing anaerobic cellulolytic bacteria: Cloning of endoglucanase genes and a β-glucosidase gene from *Clostridium cellulolyticum*. (Research Report U89008).
5. Shima, S. (1988) Cellulase Secretion from transformed *Escherichia coli* JM109. (Research Report U88055).
6. Shima, S. (1988) Cloning of *Clostridium cellobioparum* cellulase gene. (Research Report U87053).